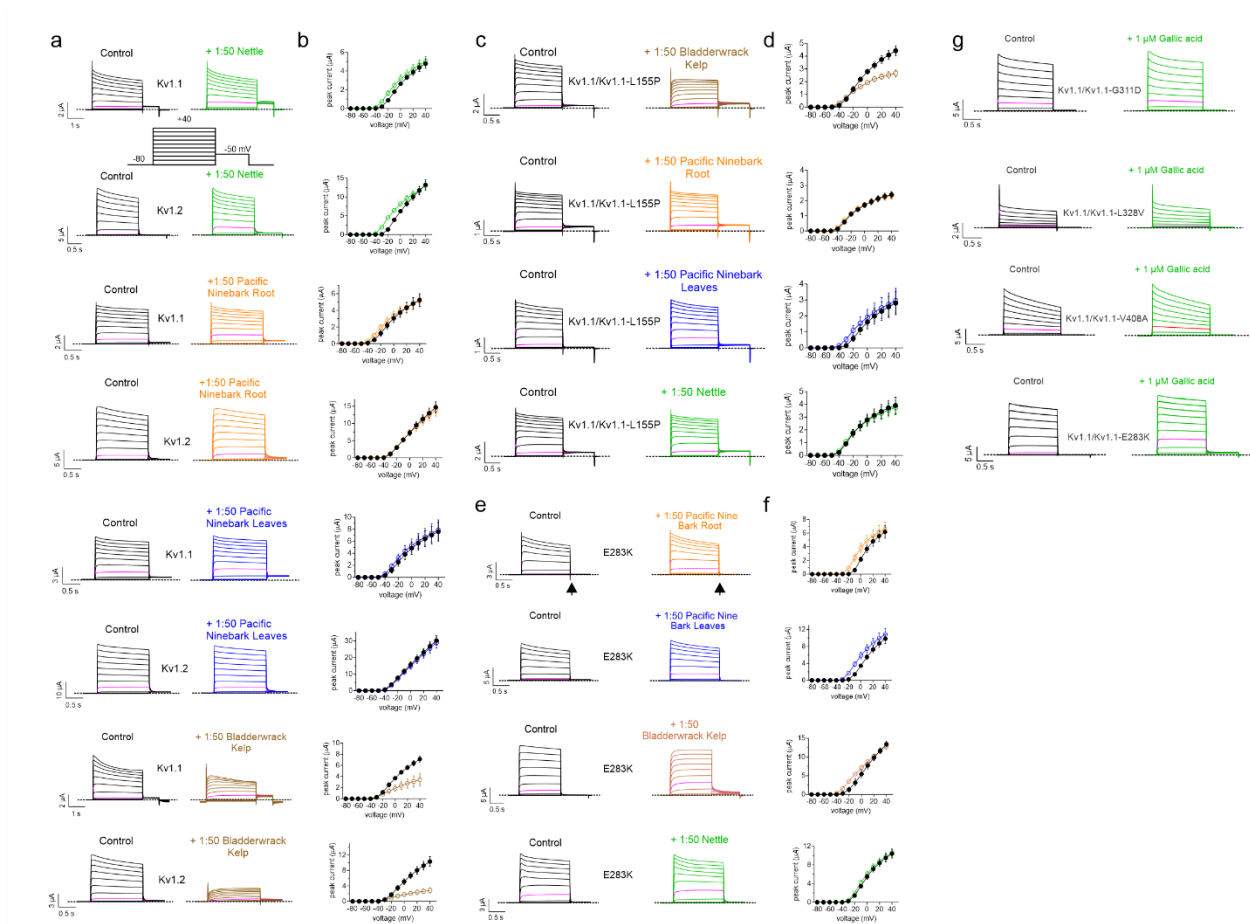


Supplementary Figure 1. Detection of gallic acid in *F. gardneri* and *P. capitatus* extracts.

Analysis of plant extracts after dilution 2- to 10-fold with MeOH was used to detect gallic acid by MS. In negative ionization mode authentic gallic acid showed molecular ion m/z 169 ($M-H^+$) and daughter ion m/z 125. *F. gardneri* and *P. capitatus* (leaves and roots) gave solutions after dilution in MeOH and were directly injected onto the LC/MS. Gallic acid was identified in extract of *F. gardneri* and *P. capitatus* root.

Preparative RPHPLC of plant extracts (panels A-D). Aqueous plant extracts were diluted with 0.1% TFA/MeOH before injection. If 1:1 dilution didn't give solutions, additional 0.1% TFA/MeOH was added until a clear solution formed.

- Injection of authentic gallic acid (2.1 mg) in 0.1% TFA/MeOH with mobile phase 15 to 100% MeOH/water (both with 0.1% TFA) gave a single peak with retention time (RT) 1.4 min with UV maximum at 270 nm.
- Injection of *F. gardneri* extract (0.25 mL diluted with 0.75 mL 0.1% TFA/MeOH) showed peaks at 1.2 min, 1.35 min and a broad peak at RT 2.1 min. Analysis of the peak with RT 1.35 min after 10-fold dilution with MeOH showed the presence of gallic acid.
- The leaf extract of *P. capitatus* gave peaks with RT 1.2 and 1.4 min but LC/MS analysis did not show gallic acid.
- Injection of the *P. capitatus* root extract (0.5 mL diluted 1:1 with 0.1% TFA/MeOH) gave a major peak with RT 1.4 min that showed gallic acid by LC/MS after 10-fold dilution with MeOH.



Supplementary Figure 2. Full voltage families and IV plots to supplement main figures.

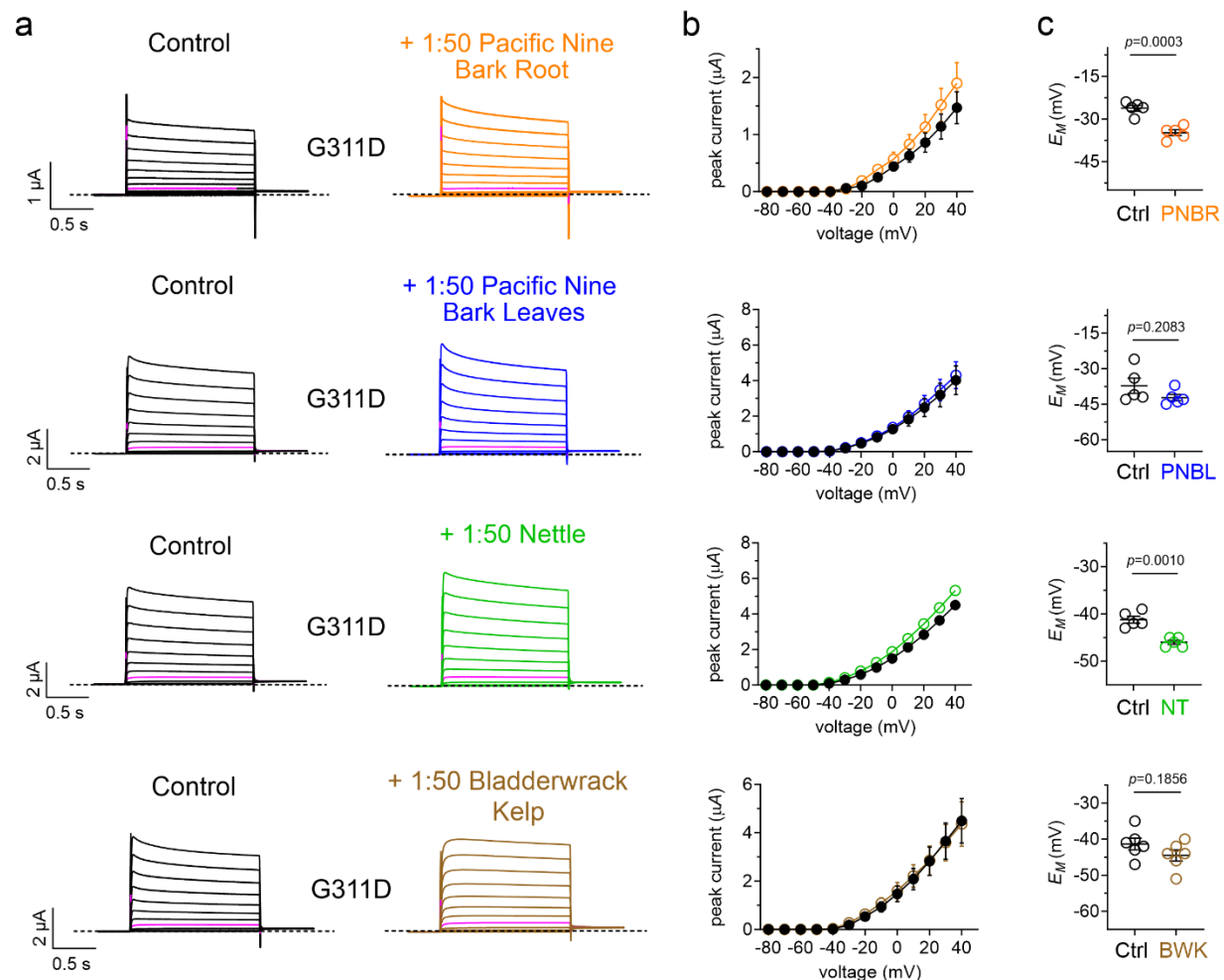
Voltage protocol as in Figure 2. Error bars indicate SEM. At least 2 batches of oocytes were used per experiment. Magenta traces indicate same-voltage traces within each pairing for ease of visual comparison.

a, b. Full voltage families (a) and IV plots (b) for groups as in Figure 1b-d; Kv1.1 nettles ($n = 5$); Kv1.2 nettles ($n = 6$); Kv1.1 pacific ninebark root ($n = 5$); Kv1.2 pacific ninebark root ($n = 5$); Kv1.1 pacific ninebark leaves ($n = 5$); Kv1.2 pacific ninebark leaves ($n = 5$); Kv1.1 bladderwrack kelp ($n = 4$); Kv1.2 bladderwrack kelp ($n = 6$). Error bars indicate SEM.

c, d. Full voltage families (c) and IV plots (d) for groups as in Figure 5b-d, $n = 5$. Error bars indicate SEM.

e, f. Full voltage families (e) and IV plots (f) for groups as in Figure 6a-c, $n = 5$. Error bars indicate SEM.

g. Full voltage families for groups as in Figure 7a-d, $n = 5$.



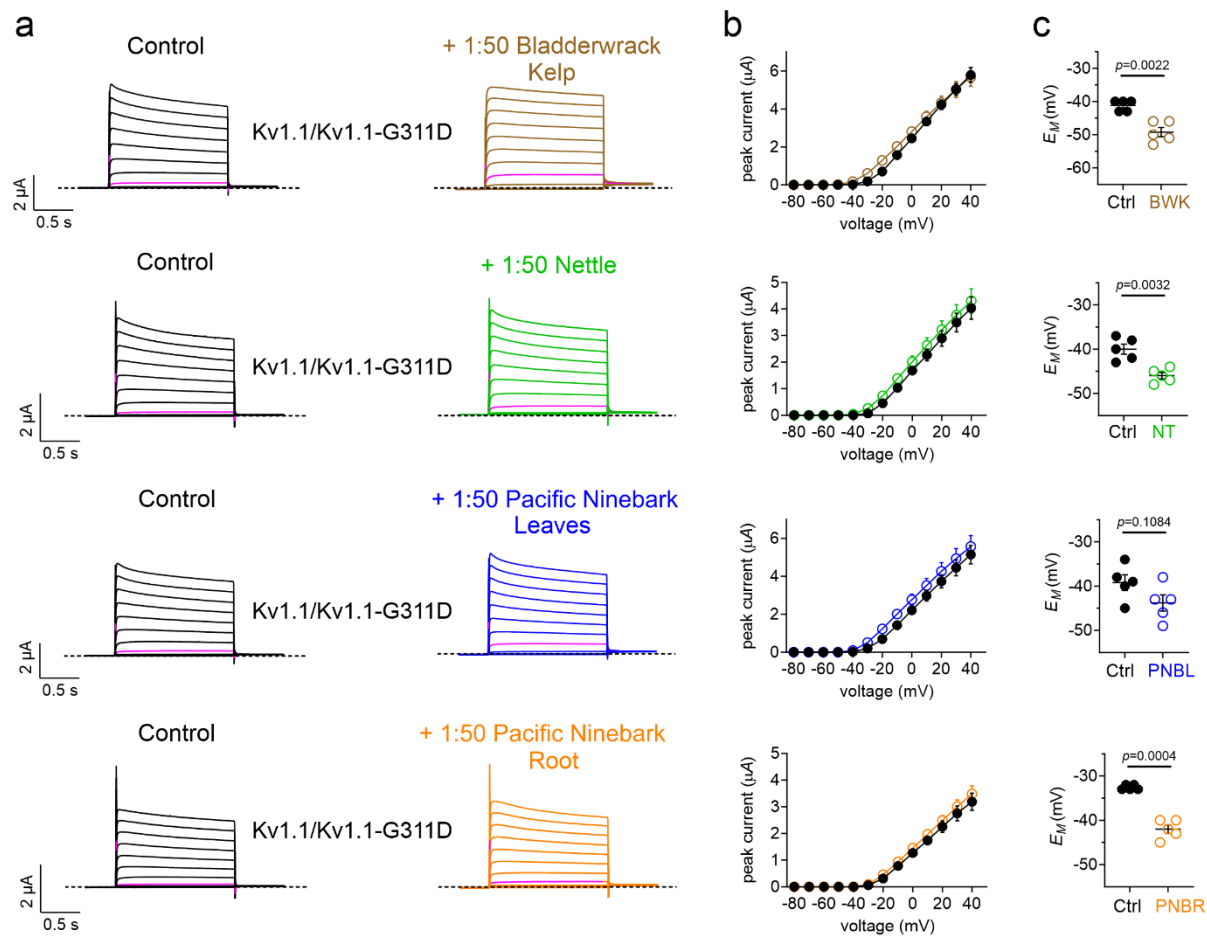
Supplementary Figure 3. Ataxia therapy extracts do not rescue the function of Kv1.1-G311D.

Voltage protocol as in Figure 2. Error bars indicate SEM; statistical analysis by two-tailed paired t-test. At least 2 batches of oocytes were used per experiment. Magenta traces indicate same-voltage traces within each pairing for ease of visual comparison.

a. Mean trace for Kv1.1-G311D in the absence (Control) and presence of plant extracts as indicated (1:50 dilution); $n = 5$.

b. Mean peak current versus voltage for Kv1.1-G311D traces as in a; $n = 5$.

c. Mean E_M for oocytes expressing Kv1.1-G311D in the absence (Control) and presence of plant extracts as in A; pacific ninebark root ($n = 5$; $p=0.0003$); pacific ninebark leaves ($n = 5$; $p=0.2083$); nettle ($n = 5$; $p=0.0010$); bladderwrack kelp ($n = 5$; $p=0.1856$).



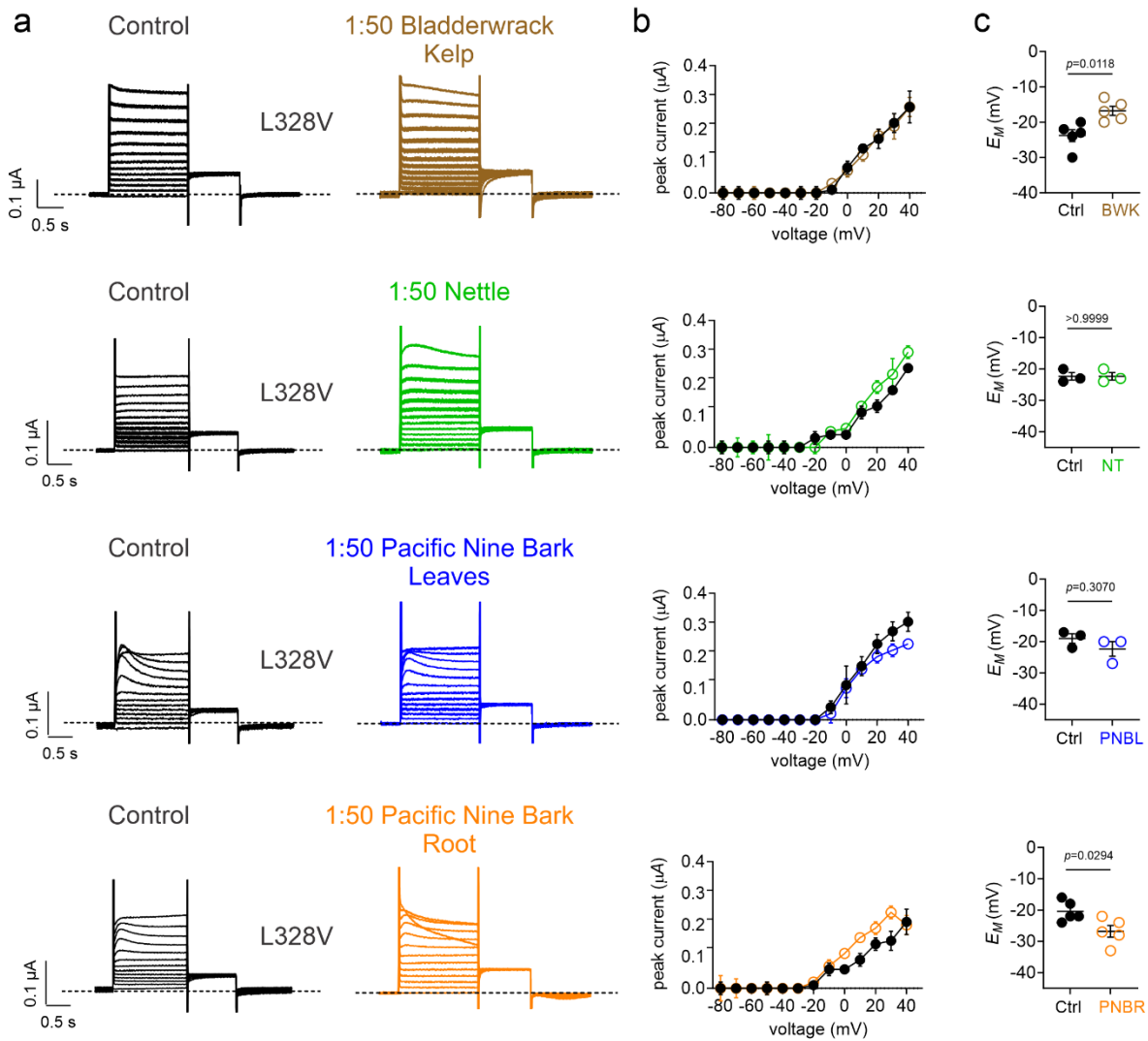
Supplementary Figure 4. Ataxia therapy extracts do not rescue the function of Kv1.1/Kv1.1-G311D.

Voltage protocol as in Figure 2. Error bars indicate SEM; statistical analysis by two-tailed paired t-test. At least 2 batches of oocytes were used per experiment. Magenta traces indicate same-voltage traces within each pairing for ease of visual comparison.

a. Mean trace for Kv1.1/Kv1.1-G311D in the absence (Control) and presence of plant extracts as indicated (1:50 dilution); $n = 5$.

b. Mean peak current versus voltage for Kv1.1/Kv1.1-G311D traces as in a; $n = 5$.

c. Mean E_M for oocytes expressing Kv1.1/Kv1.1-G311D in the absence (Control) and presence of plant extracts as in A; bladderwrack kelp ($n = 5$; $p=0.0022$); nettle ($n = 5$; $p=0.0032$); pacific ninebark leaves ($n = 5$; $p=0.1084$); pacific ninebark root ($n = 5$; $p=0.0004$).



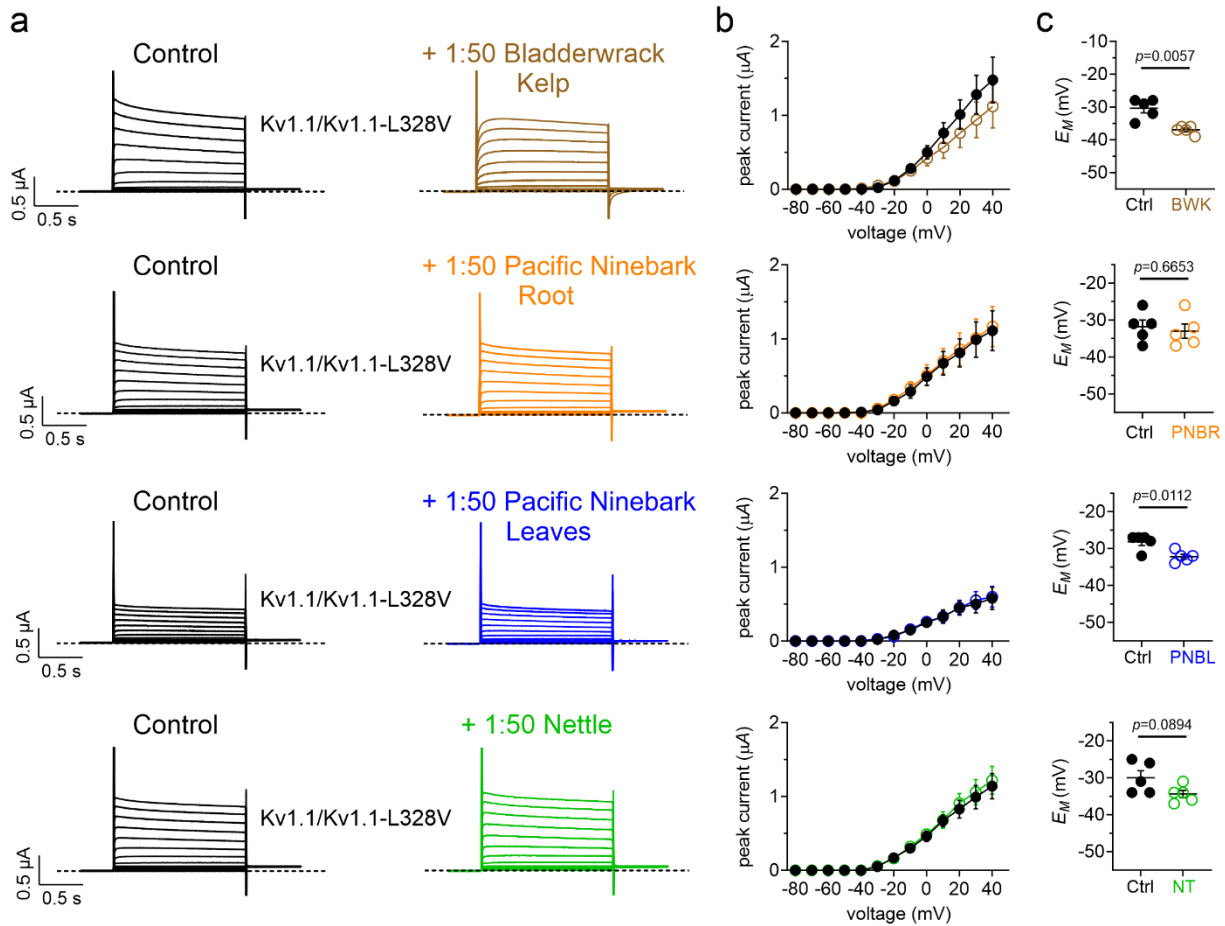
Supplementary Figure 5. Ataxia therapy plant extracts do not rescue the function of Kv1.1-L328V.

Voltage protocol as in Figure 2. Error bars indicate SEM; statistical analysis by two-tailed paired t-test. At least 2 batches of oocytes were used per experiment. Magenta traces indicate same-voltage traces within each pairing for ease of visual comparison.

a. Mean trace for Kv1.1-L328V in the absence (Control) and presence of plant extracts as indicated (1:50 dilution). bladderwrack kelp ($n = 5$); nettle ($n = 3$); pacific ninebark leaves ($n = 3$); pacific ninebark root ($n = 5$).

b. Mean peak current versus voltage for Kv1.1-L328V traces in a; bladderwrack kelp ($n = 5$); nettle ($n = 3$); pacific ninebark leaves ($n = 3$); pacific ninebark root ($n = 5$).

c. Mean E_M for oocytes expressing Kv1.1-L328V in the absence (Control) or presence of plant extracts as in a; bladderwrack kelp ($n = 5$; $p=0.0118$); nettle ($n = 3$; >0.9999); pacific ninebark leaves ($n = 3$; $p=0.3070$); pacific ninebark root ($n = 5$; $p=0.0294$).



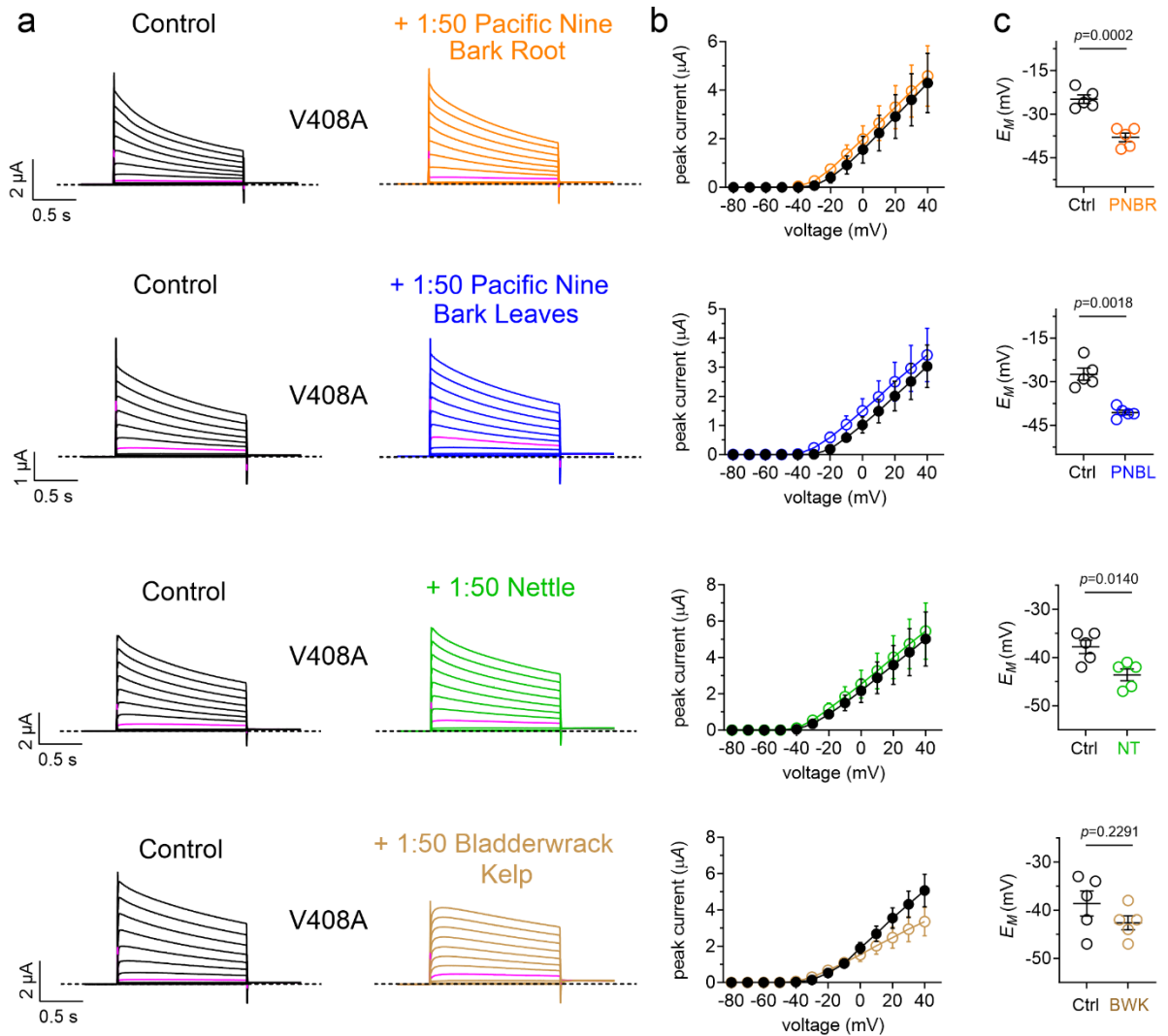
Supplementary Figure 6. Ataxia therapy plant extracts do not rescue the function of Kv1.1/Kv1.1-L328V.

Voltage protocol as in Figure 2. Error bars indicate SEM; statistical analysis by two-tailed paired t-test. At least 2 batches of oocytes were used per experiment. Magenta traces indicate same-voltage traces within each pairing for ease of visual comparison.

a. Mean trace for Kv1.1/Kv1.1-L328V in the absence (Control) and presence of plant extracts as indicated (1:50 dilution); $n = 5$.

b. Mean peak current versus voltage for Kv1.1/Kv1.1-L328V traces in a; $n = 5$.

c. Mean E_M for oocytes expressing Kv1.1/Kv1.1-L328V in the absence (Control) or presence of plant extracts as in a; bladderwrack kelp ($n = 5$; $p=0.0057$); pacific ninebark root ($n = 5$; $p=0.6653$); pacific ninebark leaves ($n = 5$; $p=0.0112$); nettle ($n = 5$; $p=0.0894$).



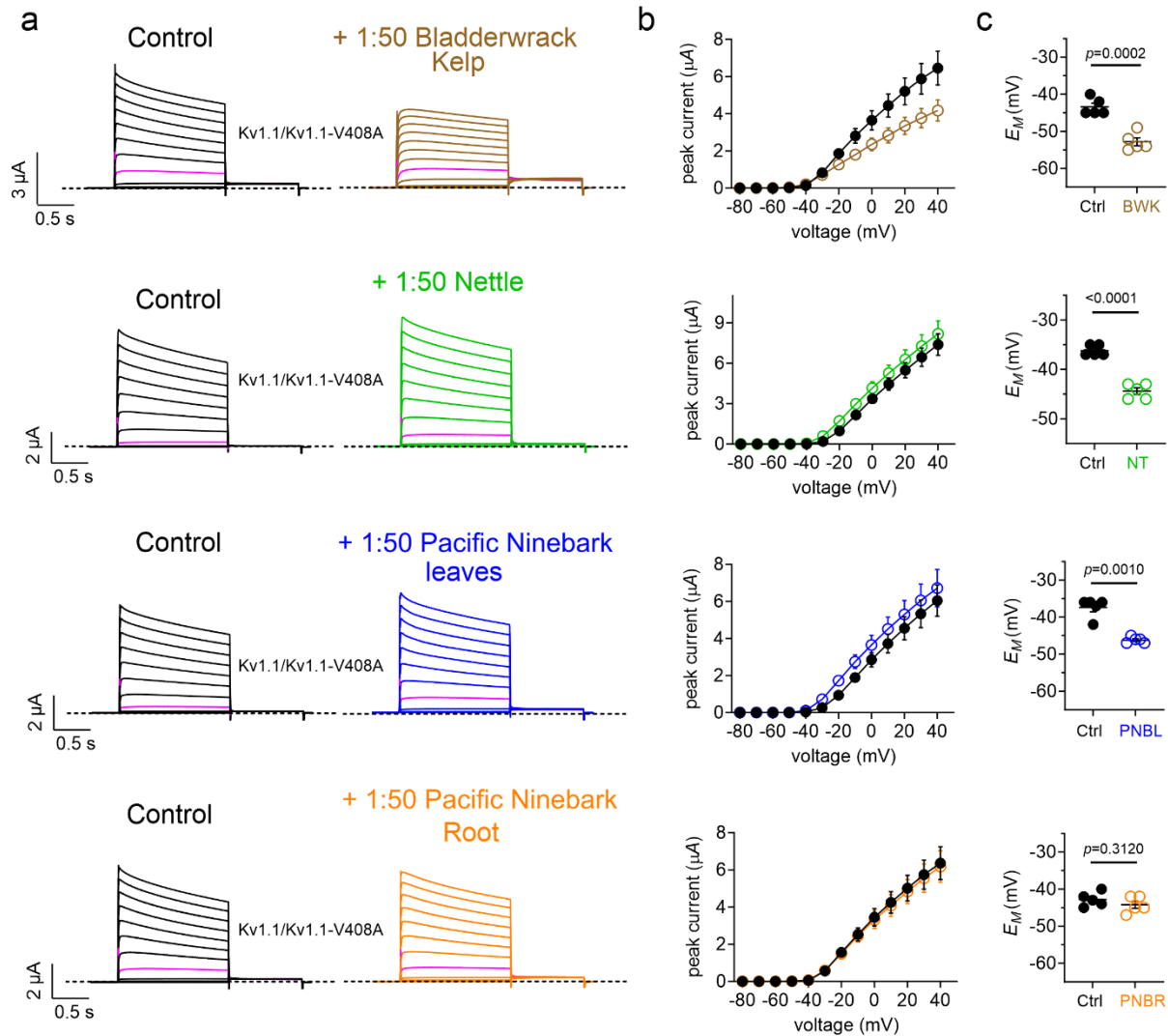
Supplementary Figure 7. Ataxia therapy plant extracts do not rescue the function of Kv1.1-V408A.

Voltage protocol as in Figure 2. Error bars indicate SEM; statistical analysis by two-tailed paired t-test. At least 2 batches of oocytes were used per experiment. Magenta traces indicate same-voltage traces within each pairing for ease of visual comparison.

a. Mean trace for Kv1.1-V408A in the absence (Control) and presence of plant extracts as indicated (1:50 dilution); $n = 5$.

b. Mean peak current versus voltage for Kv1.1-V408A traces in a; $n = 5$.

c. Mean E_M for oocytes expressing Kv1.1-V408A in the absence (Control) or presence of plant extracts as in a; pacific ninebark root ($n = 5$; $p=0.0002$); pacific ninebark leaves ($n = 5$; $p=0.0018$); nettle ($n = 5$; $p=0.0140$); bladderwrack kelp ($n = 5$; $p=0.2291$).



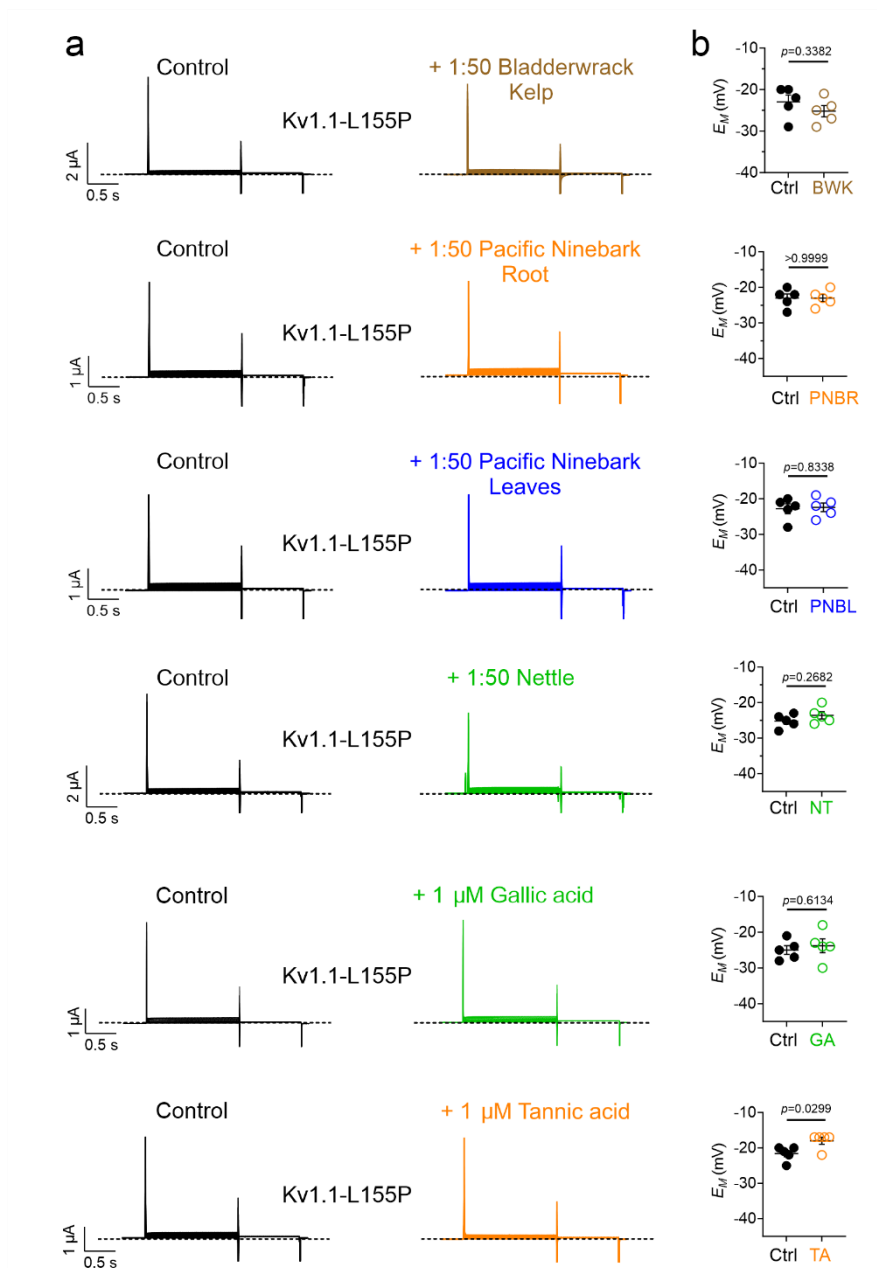
Supplementary Figure 8. Ataxia therapy plant extracts do not rescue the function of Kv1.1/Kv1.1-V408A.

Voltage protocol as in Figure 2. Error bars indicate SEM; statistical analysis by two-tailed paired t-test. At least 2 batches of oocytes were used per experiment. Magenta traces indicate same-voltage traces within each pairing for ease of visual comparison.

a. Mean trace for Kv1.1/Kv1.1-V408A in the absence (Control) and presence of plant extracts as indicated (1:50 dilution); $n = 5$.

b. Mean peak current versus voltage for Kv1.1/Kv1.1-V408A traces in a; $n = 5$.

c. Mean E_M for oocytes expressing Kv1.1/Kv1.1-V408A in the absence (Control) or presence of plant extracts as in a; bladderwrack kelp ($n = 5$; $p=0.0002$); nettle ($n = 5$; <0.0001); pacific ninebark leaves ($n = 5$; $p=0.0010$); pacific ninebark root ($n = 5$; $p=0.3120$).

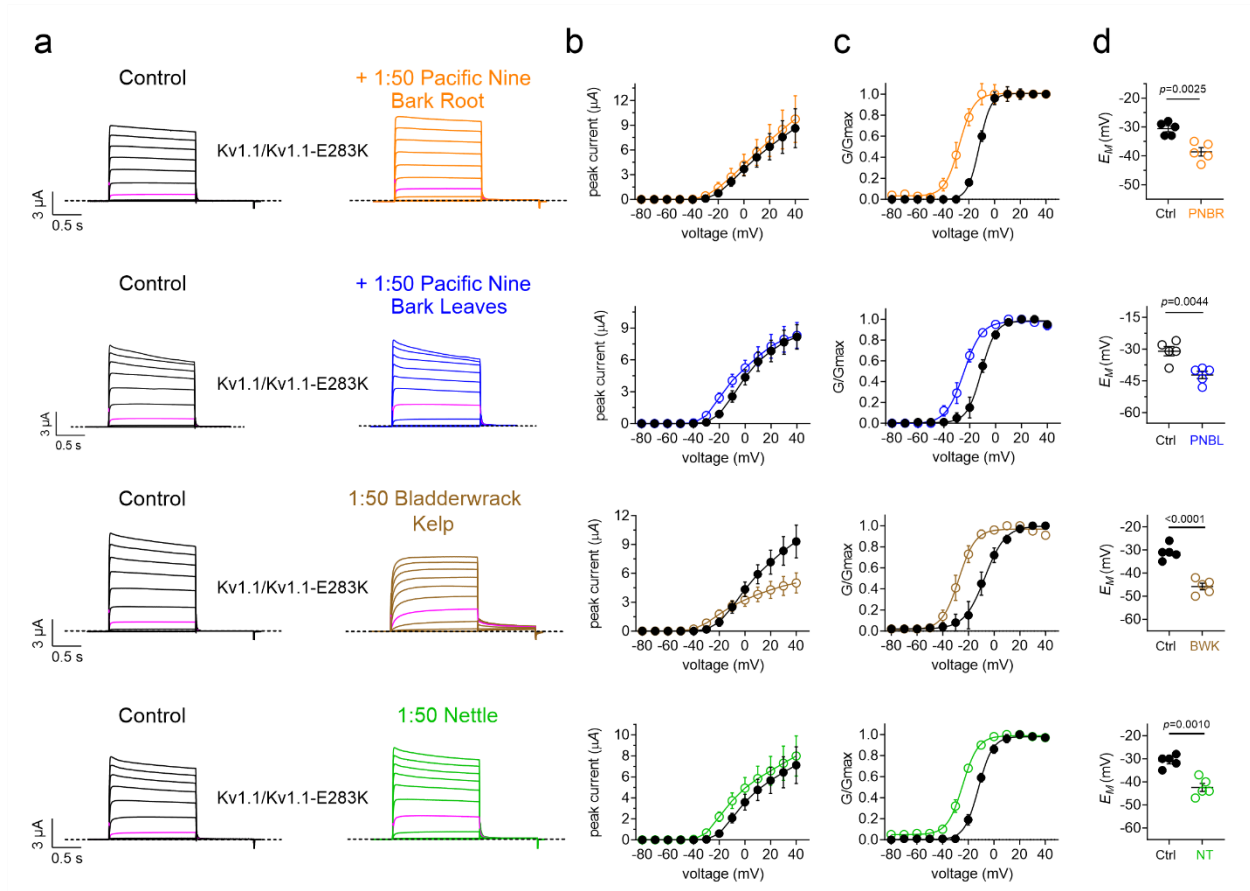


Supplementary Figure 9. Neither ataxia therapy plant extracts, nor gallic or tannic acids, rescue the function of “homozygous” Kv1.1-L155P.

Voltage protocol as in Figure 2. Error bars indicate SEM; statistical analysis by two-tailed paired t-test. At least 2 batches of oocytes were used per experiment. Magenta traces indicate same-voltage traces within each pairing for ease of visual comparison.

a. Mean trace for Kv1.1-L155P in the absence (Control) and presence of plant extracts as indicated (1:50 dilution); $n = 5$.

b. Mean E_M for oocytes expressing Kv1.1-L155P in the absence (Control) or presence of plant extracts as in a; bladderwrack kelp ($n = 5$; $p=0.3382$); pacific ninebark root ($n = 5$; >0.9999); pacific ninebark leaves ($n = 5$; $p=0.8338$); nettle ($n = 5$; $p=0.2682$); 1 μ M gallic acid ($n = 5$; $p=0.6134$); 1 μ M tannic acid ($n = 5$; $p=0.0299$).



Supplementary Figure 10. Ataxia therapy plant extracts rescue the function of Kv1.1/Kv1.1-E283K.

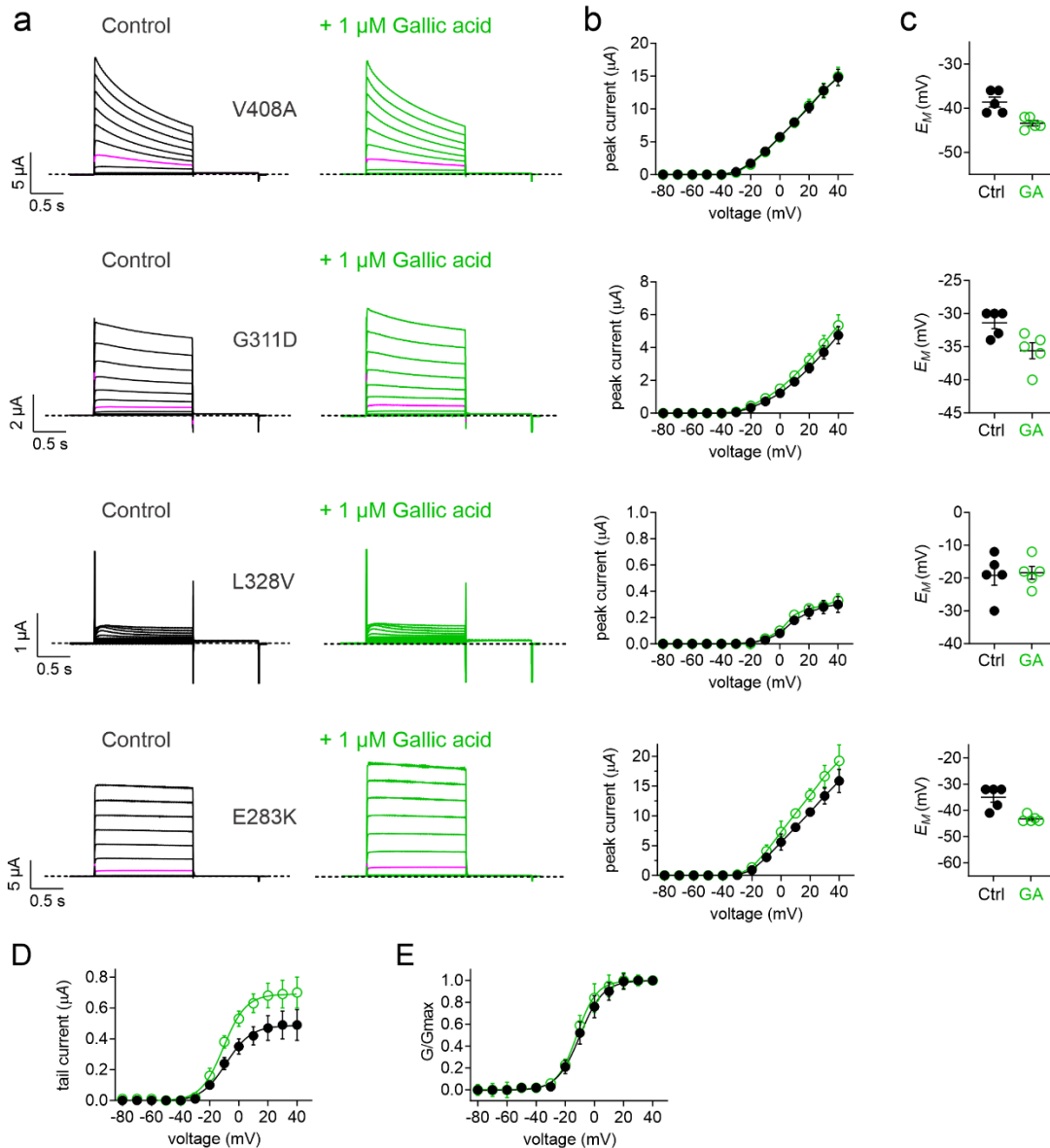
Voltage protocol as in Figure 2. Error bars indicate SEM; statistical analysis by two-tailed paired t-test. At least 2 batches of oocytes were used per experiment. Magenta traces indicate same-voltage traces within each pairing for ease of visual comparison.

a. Mean trace for Kv1.1/Kv1.1-E283K in the absence (Control) and presence of plant extracts as indicated (1:50 dilution); $n = 5$.

b. Mean peak current versus voltage for Kv1.1/Kv1.1-E283K traces in a; $n = 5$.

c. Mean G/Gmax quantified from tail current for Kv1.1/Kv1.1-E283K traces as in a; $n = 5$.

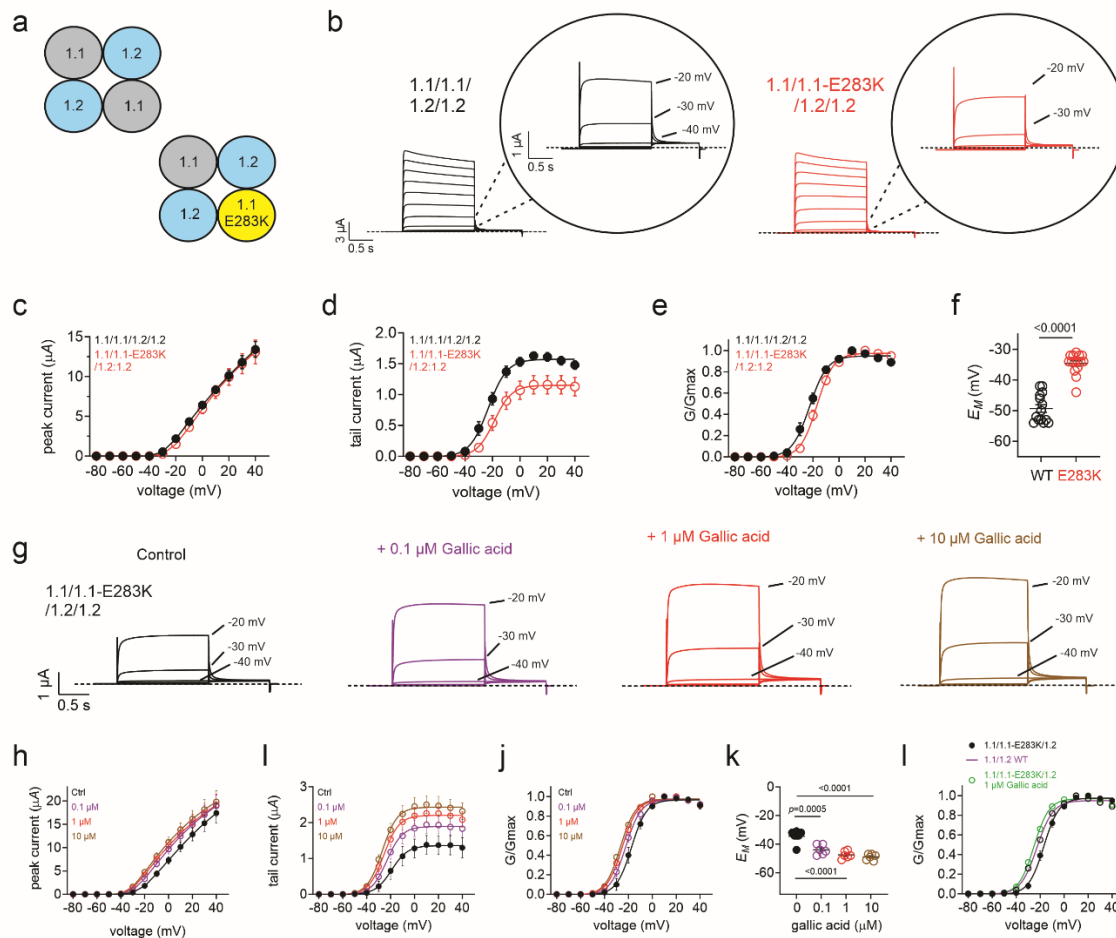
d. Mean E_M for oocytes expressing Kv1.1/Kv1.1-E283K in the absence (Control) or presence of plant extracts as in a: pacific ninebark root ($n = 5$; $p=0.0025$); pacific ninebark leaves ($n = 5$; $p=0.0044$); bladderwrack kelp ($n = 5$; <0.0001); nettle ($n = 5$; $p=0.0010$).



Supplementary Figure 11. Gallic acid (1 μM) is ineffective at rescuing the function of “homozygous” Kv1.1 ataxia mutant channels.

Voltage protocol as in Figure 2. Error bars indicate SEM; statistical analysis by two-tailed paired t-test. At least 2 batches of oocytes were used per experiment. Magenta traces indicate same-voltage traces within each pairing for ease of visual comparison.

- Mean traces for ataxia mutant Kv1.1 channels as indicated in the absence (Control) and presence of gallic acid (1 μM); $n = 5$.
- Mean peak current versus voltage for ataxia mutant Kv1.1 channels as in a; $n = 5$.
- Mean E_M for oocytes expressing ataxia mutant Kv1.1 channels in the absence (Control) or presence of plant extracts as in a; Kv1.1-V408A ($n = 5$; $p=0.0090$); Kv1.1-G311D ($n = 5$; $p=0.0248$); Kv1.1-L328V ($n = 5$; $p=0.8289$); Kv1.1-E283K ($n = 5$; $p=0.0101$).
- Mean tail current versus voltage for Kv1.1-E283K channels as in a; $n = 5$.
- Mean G/Gmax versus voltage for Kv1.1-E283K channels as in a; $n = 5$.



Supplementary Figure 12. Gallic acid rescues the function of EA1-linked E283K heteromeric Kv1.1-Kv1.2 channels.

Voltage protocol as in Figure 2. Error bars indicate SEM; statistical analysis by two-tailed paired t-test or One-Way ANOVA. At least 2 batches of oocytes were used per experiment.

a. cartoon representing the ratios of Kv1.x cRNA injected into each oocyte.

b. Mean traces for heteromeric wild-type (left; $n = 14$) and E283K (right; $n = 15$) Kv1.1/Kv1.2 channels expressed in oocytes; scale bars lower left. Bubbles indicate vertical scale expanded region to show reduced current in mutant channels at mildly depolarized potentials.

c-e. Mean peak, tail and normalized tail (G/G_{max}) currents versus voltage for heteromeric wild-type (left; $n = 14$) and E283K (right; $n = 15$) Kv1.1/Kv1.2 channels.

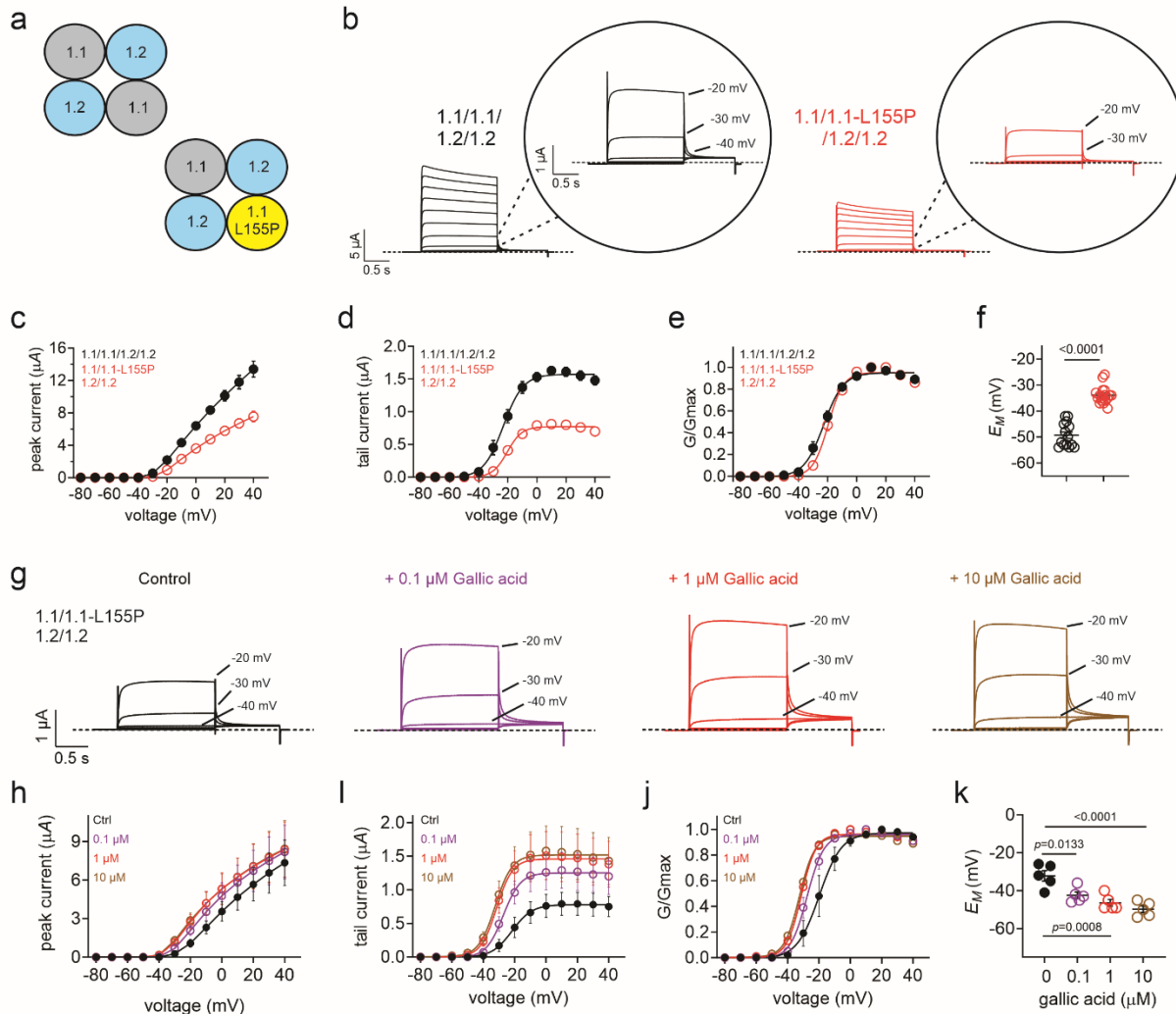
f. Mean E_M for oocytes expressing heteromeric wild-type (left; $n = 14$) and E283K (right; $n = 15$) Kv1.1/Kv1.2 channels (<0.0001).

g. Mean current traces for Kv1.1/Kv1.1-E283K/Kv1.2 channels in the absence or presence of gallic acid doses as indicated ($n = 6$).

h-j. Mean peak, tail, and normalized (G/G_{max}) currents versus voltage for channels as in g; $n = 6$.

k. Mean E_M for oocytes expressing Kv1.1/Kv1.1-E283K/Kv1.2 channels in: 0.1 μ M gallic acid ($n = 6$; $p = 0.0005$); 1 μ M gallic acid ($n = 6$; <0.0001); 10 μ M gallic acid ($n = 6$; <0.0001).

l. Comparison of mean normalized tail currents (G/G_{max}) showing that gallic acid (1 μ M) returns mutant E283K Kv1.1/Kv1.2 ($n = 6$) channel voltage dependence to match that heteromeric wild type ($n = 14$).



Supplementary Figure 13. Gallic acid rescues the function of EA1-linked L155P heteromeric Kv1.1-Kv1.2 channels.

Voltage protocol as in Figure 2. Error bars indicate SEM; statistical analysis by two-tailed paired t-test or One-Way ANOVA. At least 2 batches of oocytes were used per experiment.

a. cartoon representing the ratios of Kv1.x cRNA injected into each oocyte.

b. Mean traces for heteromeric wild-type (left; $n = 14$) and L155P (right; $n = 18$) Kv1.1/Kv1.2 channels expressed in oocytes; scale bars lower left. Bubbles indicate vertical scale expanded region to show reduced current in mutant channels at mildly depolarized potentials.

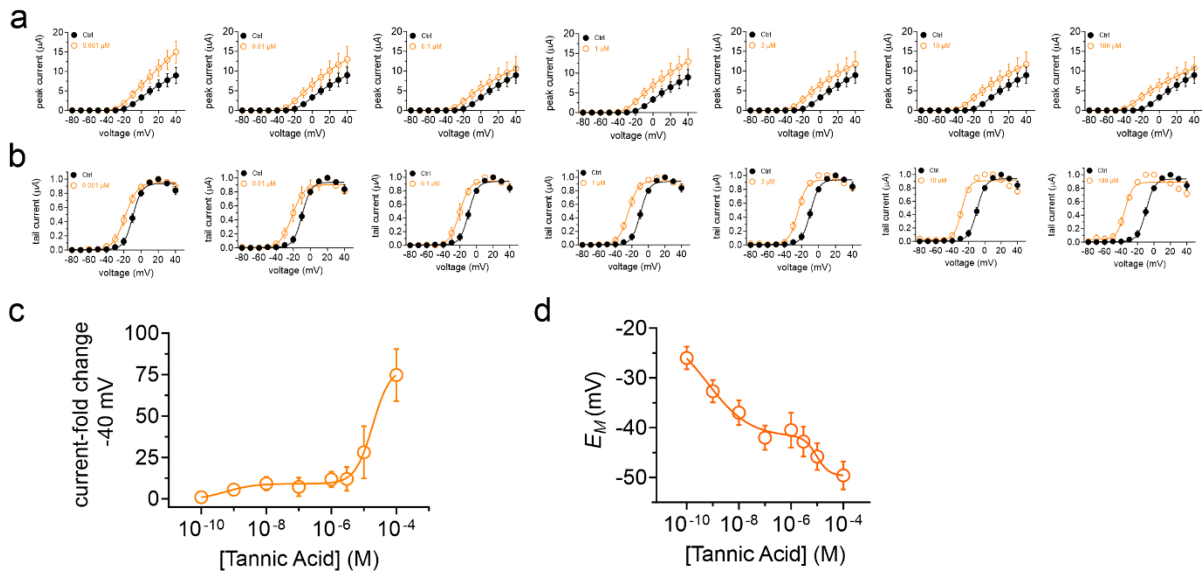
c-e. Mean peak, tail, and normalized tail (G/G_{max}) currents versus voltage for heteromeric wild-type (left; $n = 14$) and L155P (right; $n = 18$) Kv1.1/Kv1.2 channels as in b.

f. Mean E_M for oocytes expressing heteromeric wild-type (left; $n = 14$) and L155P (right; $n = 18$) Kv1.1/Kv1.2 channels as in b (<0.0001).

g. Mean current traces for Kv1.1/Kv1.1-L155P/Kv1.2 channels in the absence or presence of gallic acid doses as indicated ($n = 5$).

h-j. Mean peak, tail, and normalized (G/G_{max}) currents versus voltage for channels as in g; $n = 5$.

k. Mean E_M for oocytes expressing Kv1.1/Kv1.1-L155P/Kv1.2 channels in; 0.1 μ M gallic acid ($n = 5$; $p=0.0133$); 1 μ M gallic acid ($n = 5$; $p=0.0008$); 10 μ M gallic acid ($n = 5$; <0.0001).



Supplementary Figure 14. Dose response for tannic acid effects on Kv1.1-E283K.

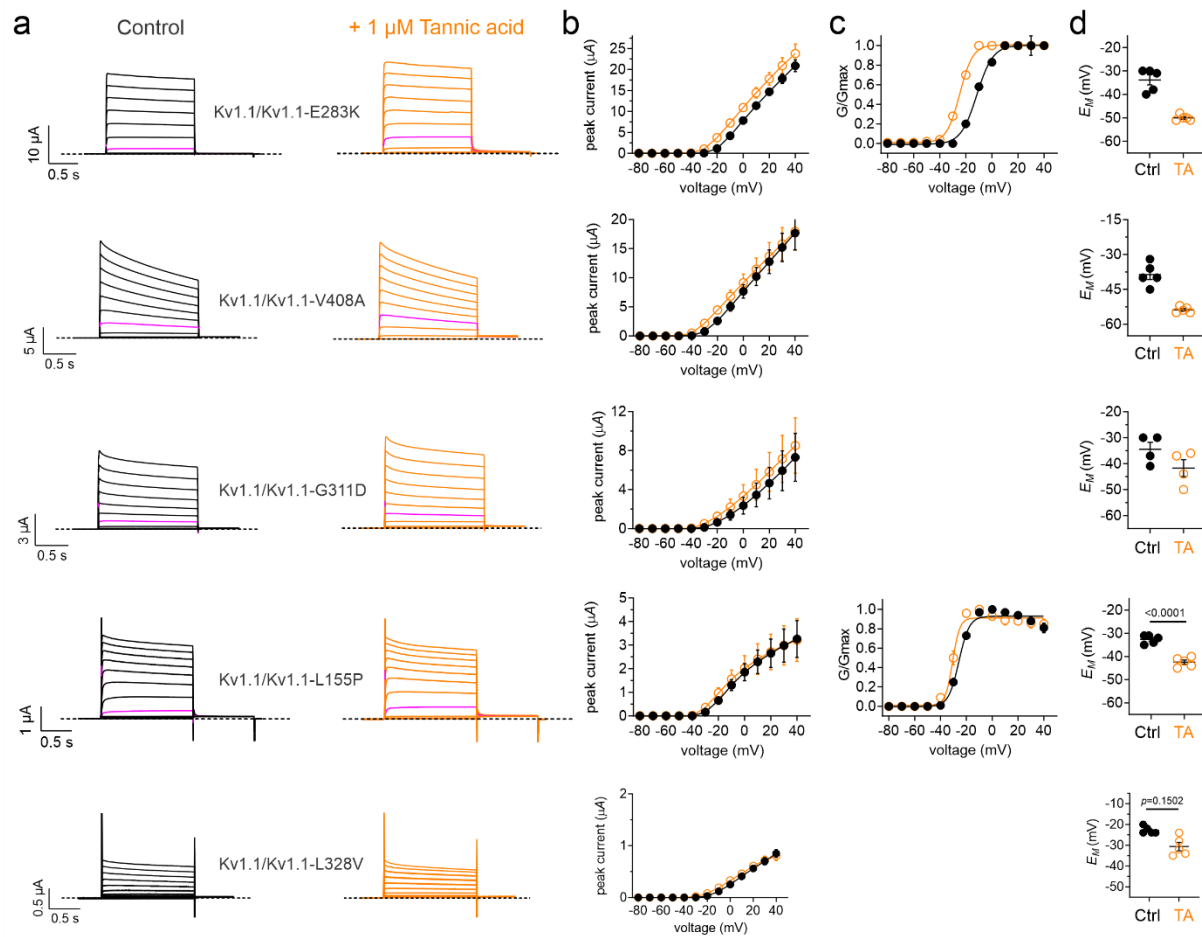
Voltage protocol as in Figure 2. Error bars indicate SEM. At least 2 batches of oocytes were used per experiment.

a. Mean peak current versus voltage for Kv1.1-E283K in the absence (Control) or presence of tannic acid concentrations: 0.001 μM ($n = 4$); 0.01 μM ($n = 4$); 0.1 μM ($n = 6$); 1 μM ($n = 4$); 3 μM ($n = 6$); 10 μM ($n = 6$); 100 μM ($n = 6$).

b. Mean G/Gmax versus voltage for Kv1.1-E283K in the absence (Control) or presence of tannic acid concentrations as indicated: 0.001 μM ($n = 4$); 0.01 μM ($n = 4$); 0.1 μM ($n = 6$); 1 μM ($n = 4$); 3 μM ($n = 6$); 10 μM ($n = 6$); 100 μM ($n = 6$).

c. Dose response for tannic acid effects at -40 mV on Kv1.1-E283K calculated from graphs as in A. 0.001 μM ($n = 3$); 0.01 μM ($n = 4$); 0.1 μM ($n = 6$); 1 μM ($n = 4$); 3 μM ($n = 6$); 10 μM ($n = 6$); 100 μM ($n = 6$).

d. Dose response for tannic acid effects on E_M of oocytes expressing Kv1.1-E283K, calculated from as in a: 0.001 μM ($n = 4$); 0.01 μM ($n = 4$); 0.1 μM ($n = 6$); 1 μM ($n = 4$); 3 μM ($n = 6$); 10 μM ($n = 6$); 100 μM ($n = 6$).



Supplementary Figure 15. Tannic acid (1 μ M) enhances Kv1.1/Kv1.1-E283K but no other mixed wild-type/ataxia mutant Kv1.1 channels.

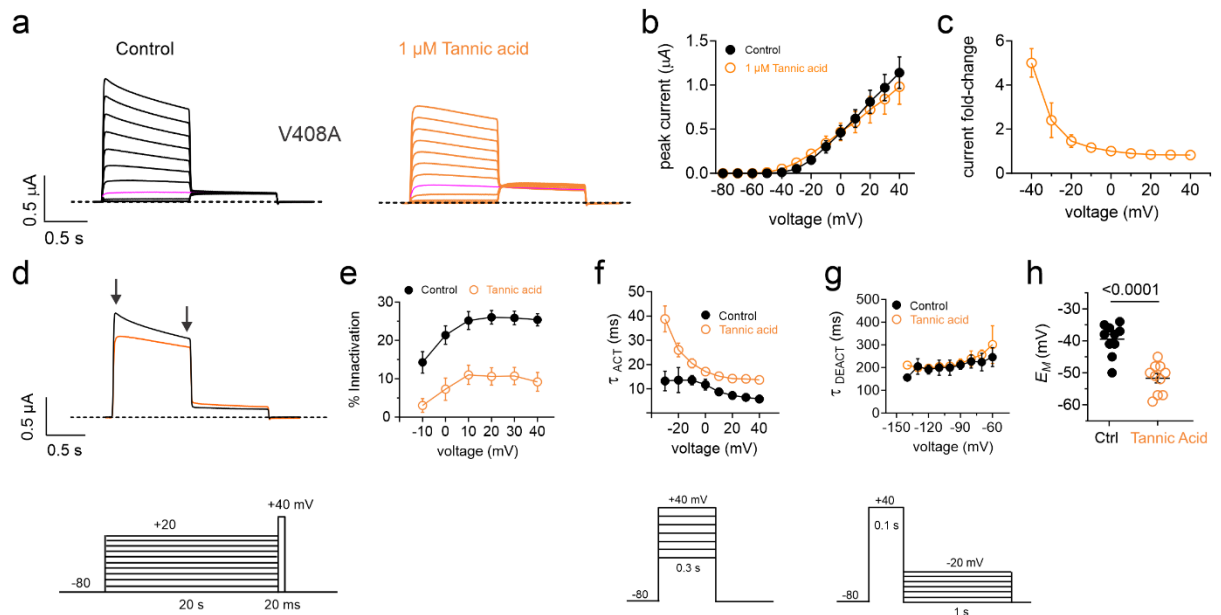
Voltage protocol as in Figure 2. Error bars indicate SEM; statistical analysis by two-tailed paired t-test. At least 2 batches of oocytes were used per experiment. Magenta traces indicate same-voltage traces within each pairing for ease of visual comparison.

a. Mean trace for heteromeric channels as indicated in the absence (Control) and presence of tannic acid (1 μ M): Kv1.1/Kv1.1-E283K ($n = 5$); Kv1.1/Kv1.1-V408A ($n = 5$); Kv1.1/Kv1.1-G311D ($n = 4$); Kv1.1/Kv1.1-L155P ($n = 5$); Kv1.1/Kv1.1-L328V ($n = 5$).

b. Mean peak currents versus voltage for traces as in a: Kv1.1/Kv1.1-E283K ($n = 5$); Kv1.1/Kv1.1-V408A ($n = 5$); Kv1.1/Kv1.1-G311D ($n = 4$); Kv1.1/Kv1.1-L155P ($n = 5$); Kv1.1/Kv1.1-L328V ($n = 5$).

c. Mean G/Gmax versus voltage for traces as in a: Kv1.1/Kv1.1-E283K ($n = 5$); Kv1.1/Kv1.1-V408A ($n = 5$); Kv1.1/Kv1.1-G311D ($n = 4$); Kv1.1/Kv1.1-L155P ($n = 5$); Kv1.1/Kv1.1-L328V ($n = 5$). Graphs omitted where tail currents were too small to quantify.

d. Mean E_M for oocytes expressing channels as in a in the absence (Control) or presence of tannic acid (1 μ M) Kv1.1/Kv1.1-E283K ($n = 5$; $p=0.0012$); Kv1.1/Kv1.1-V408A ($n = 5$; $p=0.0016$); Kv1.1/Kv1.1-G311D ($n = 4$; $p=0.1413$); Kv1.1/Kv1.1-L155P ($n = 5$; <0.0001); Kv1.1/Kv1.1-L328V ($n = 5$; $p=0.1502$).



Supplementary Figure 16. Tannic acid (1 μM) effects on Kv1.1-V408A channels.

Error bars indicate SEM; statistical analysis by two-tailed paired t-test. At least 2 batches of oocytes were used per experiment. Magenta traces indicate same-voltage traces within each pairing for ease of visual comparison.

a. Mean trace for homomeric Kv1.1-V408A channels in the absence (Control) and presence of tannic acid (1 μM); $n = 10$; Voltage protocols as in Figure 2.

b. Mean peak current versus voltage for traces as in a; $n = 10$.

c. Current fold change induced by (1 μM) tannic acid versus voltage for traces as in a; $n = 10$.

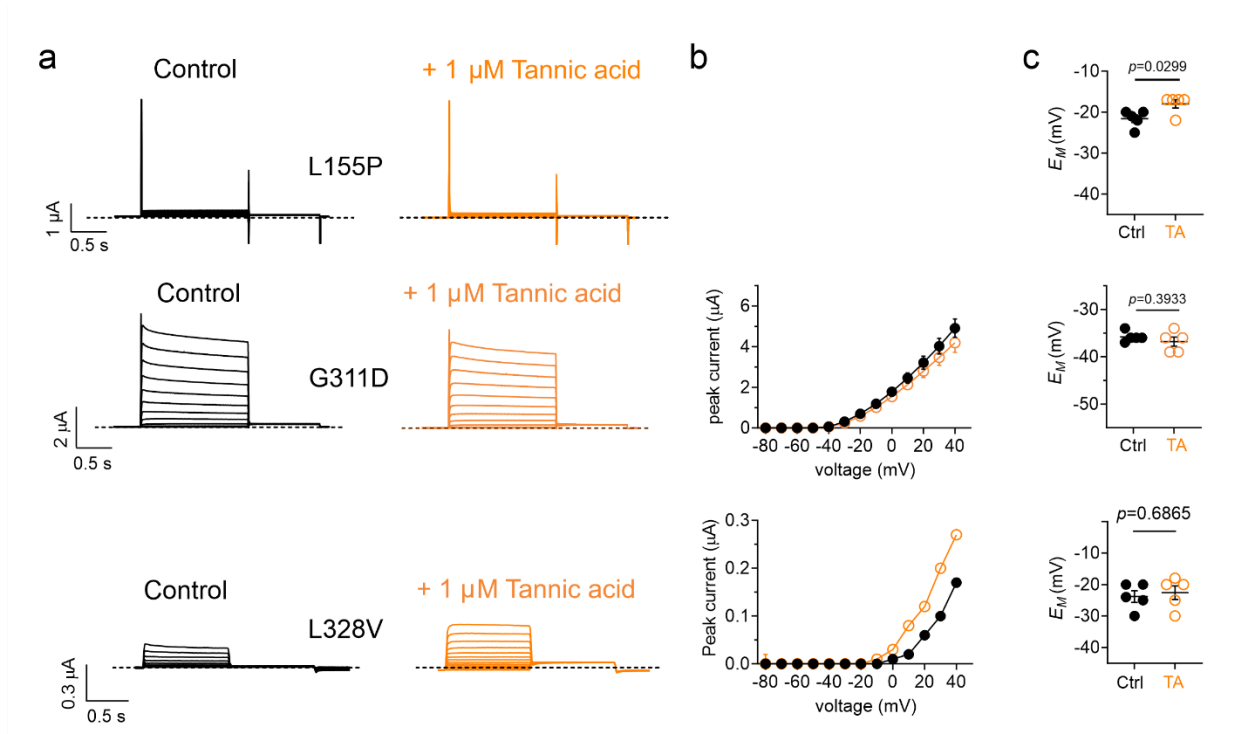
d. Mean traces showing effects of tannic acid (1 μM) on Kv1.1-V408A inactivation (between the two vertical bars) quantified using the voltage protocol shown (lower inset); $n = 10$.

e. Effects of tannic acid (1 μM) on % inactivation quantified as in d; $n = 10$.

f. Mean activation rate (τ_{ACT}) versus voltage for Kv1.1-V408A in bath solution (black) versus tannic acid (1 μM) (brown), quantified using the voltage protocol shown (lower inset); $n = 10$.

g. Mean deactivation rate (τ_{DEACT}) versus voltage for Kv1.1-V408A in bath solution (black) versus tannic acid (1 μM) (brown), quantified using the voltage protocol shown (lower inset); $n = 8$.

h. Mean E_M for oocytes expressing channels as in a in the absence (Control) or presence of tannic acid (1 μM); ($n = 10$; <0.0001).



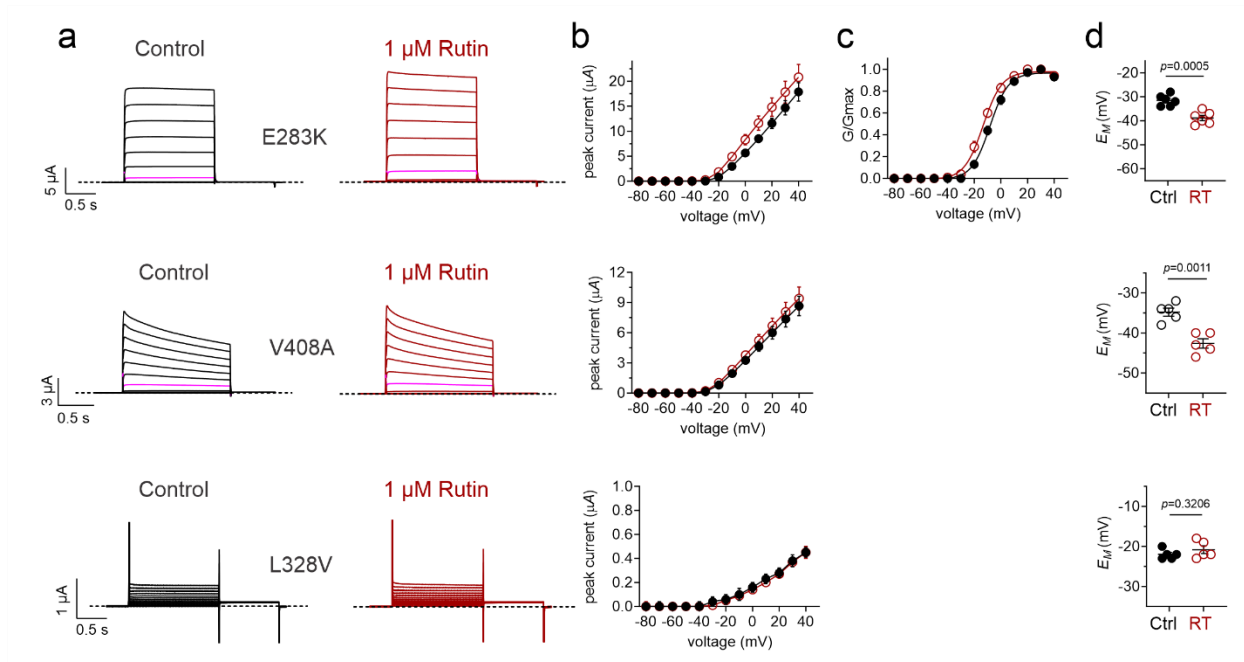
Supplementary Figure 17. Tannic acid (1 μ M) does not rescue 100% mutant L155P, G311D or L328V Kv1.1 activity.

Voltage protocol as in Figure 2. Error bars indicate SEM; statistical analysis by two-tailed paired t-test. At least 2 batches of oocytes were used per experiment.

a. Mean trace for channel as indicated in the absence (Control) and presence of tannic acid (1 μ M); $n = 5$.

b. Mean peak current versus voltage for traces as in a; $n = 5$.

c. Mean E_M for oocytes expressing channels as in a in the absence (Control) and presence Tannic acid (1 μ M): Kv1.1-L155P ($n = 5$; $p=0.0299$); Kv1.1-G311D ($n = 5$; $p=0.3933$); Kv1.1-L328V ($n = 5$; $p=0.6865$).



Supplementary Figure 18. Rutin (1 μM) is ineffective at enhancing ataxia mutant Kv1.1 channel activity.

Voltage protocol as in Figure 2. Error bars indicate SEM; statistical analysis by two-tailed paired t-test. At least 2 batches of oocytes were used per experiment. Magenta traces indicate same-voltage traces within each pairing for ease of visual comparison.

a. Mean trace for channels as indicated in the absence (Control) and presence of rutin (1 μM): Kv1.1-E283K ($n = 6$); Kv1.1-V408A ($n = 5$); Kv1.1-L328V ($n = 5$).

b. Mean peak current versus voltage for traces as in a: Kv1.1-E283K ($n = 6$); Kv1.1-V408A ($n = 5$); Kv1.1-L328V ($n = 5$).

c. Mean G/G_{max} versus voltage for traces as in a: Kv1.1-E283K ($n = 6$); Kv1.1-V408A ($n = 5$); Kv1.1-L328V ($n = 5$). Graphs omitted where tail currents were too small to quantify.

d. Mean E_M for oocytes expressing channels as in a in the absence (Control) and presence of rutin (1 μM); Kv1.1-E283K ($n = 6$; $p=0.0005$); Kv1.1-V408A ($n = 5$; $p=0.0011$); Kv1.1-L328V ($n = 5$; $p=0.3206$).

Supplementary Data – values and statistics tabulated by figure number.

Figure 1

	V_{0.5} Normalized tail current (mV)	Slope (mV)	E_M (mV)
Control	-27.13 ± 1.47	5.56 ± 1.47	-39.80 ± 1.73
1:50 Nettle	-42.46 ± 1.94 (<i>p</i> =0.0003; <i>n</i> =5)	7.46 ± 1.69 (<i>p</i> =0.4214; <i>n</i> =5)	-45.40 ± 1.23 (<i>p</i> =0.0211; <i>n</i> =5)

Statistics versus Kv1.1 in absence of Nettle. Values indicate mean ± SEM.

	V_{0.5} Normalized tail current (mV)	Slope (mV)	E_M (mV)
Control	-26.02 ± 1.15	5.93 ± 0.98	-41.20 ± 1.40
1:50 Pacific Ninebark Root	-38.85 ± 1.31 (<0.0001; <i>n</i> =5)	4.48 ± 1.25 (<i>p</i> =0.3895; <i>n</i> =5)	-51.80 ± 1.97 (<i>p</i> =0.0028; <i>n</i> =5)

Statistics versus Kv1.1 in absence of Pacific Ninebark Root. Values indicate mean ± SEM.

	V_{0.5} Normalized tail current (mV)	Slope (mV)	E_M (mV)
Control	-25.42 ± 1.29	6.45 ± 1.12	-47.40 ± 2.00
1:50 Pacific Ninebark Leaves	-36.56 ± 1.19 (<i>p</i> =0.0002; <i>n</i> =5)	6.39 ± 1.04 (<i>p</i> =0.9590; <i>n</i> =5)	-53.60 ± 1.34 (<i>p</i> =0.0201; <i>n</i> =5)

Statistics versus Kv1.1 in absence of Pacific Ninebark Leaves. Values indicate mean ± SEM.

	V_{0.5} Normalized tail current (mV)	Slope (mV)	E_M (mV)
Control	-24.39 ± 0.77	6.76 ± 0.67	-36.80 ± 1.36
1:50 Bladderwrack Kelp	-41.82 ± 1.02 (<0.0001; <i>n</i> =5)	3.30 ± 1.04 (<i>p</i> =0.0273; <i>n</i> =5)	-44.80 ± 1.34 (<i>p</i> =0.0097; <i>n</i> =5)

Statistics versus Kv1.1 in absence of Bladderwrack Kelp. Values indicate mean ± SEM.

	V_{0.5} Normalized tail current (mV)	Slope (mV)	E_M (mV)
Control	-16.86 ± 0.38	4.97 ± 0.33	-30.50 ± 1.41
1:50 Nettle	-30.23 ± 0.71 (<0.0001; <i>n</i> =5)	4.36 ± 0.71 (<i>p</i> =0.4673; <i>n</i> =5)	-44.67 ± 2.41 (<i>p</i> =0.0007; <i>n</i> =5)

Statistics versus Kv1.2 in absence of Nettle. Values indicate mean ± SEM.

	V_{0.5} Normalized tail current (mV)	Slope (mV)	E_M (mV)
Control	-25.04 ± 0.45	6.45 ± 0.39	-39.60 ± 1.56
1:50 Pacific Ninebark Root	-27.28 ± 0.58 (<i>p</i> =0.0170; <i>n</i> =5)	6.13 ± 0.50 (<i>p</i> =0.6282; <i>n</i> =5)	-42.60 ± 0.61 (<i>p</i> =0.0915; <i>n</i> =5)

Statistics versus Kv1.2 in absence of Pacific Ninebark Root. Values indicate mean ± SEM.

	V_{0.5} Normalized tail current (mV)	Slope (mV)	E_M (mV)
Control	-28.60 ± 0.40	7.46 ± 0.35	-48.00 ± 2.05
1:50 Pacific Ninebark Leaves	-28.31 ± 0.46 (<i>p</i> =0.6472; n=5)	7.17 ± 0.41 (<i>p</i> =0.6056; n=5)	-46.20 ± 1.80 (<i>p</i> =0.4156; n=5)

Statistics versus Kv1.2 in absence of Pacific Ninebark Leaves. Values indicate mean ± SEM.

	V_{0.5} Normalized tail current (mV)	Slope (mV)	E_M (mV)
Control	-19.95 ± 0.83	6.40 ± 0.73	-33.67 ± 1.20
1:50 Bladderwrack Kelp	-34.48 ± 2.13 (<i>p</i> =0.0005; n=6)	6.86 ± 1.86 (<i>p</i> =0.83; n=6)	-48.67 ± 0.7 (<i>p</i> =0.0009; n=6)

Statistics versus Kv1.2 in absence of Bladderwrack Kelp. Values indicate mean ± SEM.

Figure 2

	V_{0.5} Normalized tail current (mV)	Slope (mV)	E_M (mV)
Control	-11.67 ± 0.96	6.56 ± 0.84	-29.33 ± 1.92
100 µM Catechin Hydrate	-14.70 ± 0.44 (<i>p</i> =0.0242; <i>n</i> =6)	5.83 ± 0.38 (<i>p</i> =0.4549; <i>n</i> =6)	-31.33 ± 2.31 (<i>p</i> =0.3238; <i>n</i> =6)

Statistics versus Kv1.1 in absence of catechin hydrate. Values indicate mean ± SEM.

	V_{0.5} Normalized tail current (mV)	Slope (mV)	E_M (mV)
Control	-11.55 ± 1.55	9.57 ± 1.40	-36.20 ± 0.78
100 µM Gallic acid	-37.17 ± 2.59 (<0.0001; <i>n</i> =5)	12.10 ± 2.32 (<i>p</i> =0.3835; <i>n</i> =5)	-49.60 ± 0.91 (<i>p</i> =0.0002; <i>n</i> =5)

Statistics versus Kv1.1 in absence of Gallic acid. Values indicate mean ± SEM.

	V_{0.5} Normalized tail current (mV)	Slope (mV)	E_M (mV)
Control	-22.35 ± 0.42	6.25 ± 0.56	-40.00 ± 2.53
100 µM Cytisine	-23.96 ± 1.37 (<i>p</i> =0.3148; <i>n</i> =5)	8.15 ± 1.31 (<i>p</i> =0.2357; <i>n</i> =5)	-41.60 ± 1.77 (<i>p</i> =0.3949; <i>n</i> =5)

Statistics versus Kv1.1 in absence of Cytisine. Values indicate mean ± SEM.

	V_{0.5} Normalized tail current (mV)	Slope (mV)	E_M (mV)
Control	-21.99 ± 0.91	9.43 ± 0.92	-45.75 ± 1.20
100 µM Kaempferol	-22.40 ± 0.84 (<i>p</i> =0.8458; <i>n</i> =5)	8.68 ± 0.87 (<i>p</i> =0.5700; <i>n</i> =5)	-45.25 ± 1.83 (<i>p</i> =0.6997; <i>n</i> =5)

Statistics versus Kv1.1 in absence of Kaempferol. Values indicate mean ± SEM.

	V_{0.5} Normalized tail current (mV)	Slope (mV)	E_M (mV)
Control	-25.70 ± 1.83	6.61 ± 1.10	-43.75 ± 3.30
100 µM Quercetin	-25.12 ± 1.81 (<i>p</i> =0.8292; <i>n</i> =4)	6.00 ± 1.06 (>0.9999; <i>n</i> =4)	-43.50 ± 1.46 (<i>p</i> =0.9438; <i>n</i> =4)

Statistics versus Kv1.1 in absence of Quercetin. Values indicate mean ± SEM.

	V_{0.5} Normalized tail current (mV)	Slope (mV)	E_M (mV)
Control	-22.70 ± 1.29	8.58 ± 1.28	-44.50 ± 1.73
100 µM Rutin	-31.59 ± 3.22 (<i>p</i> =0.0634; <i>n</i> =4)	11.79 ± 2.88 (<i>p</i> =0.3642; <i>n</i> =4)	-56.75 ± 0.28 (<i>p</i> =0.0065; <i>n</i> =4)

Statistics versus Kv1.1 in absence of Rutin. Values indicate mean ± SEM.

	V_{0.5} Normalized tail current (mV)	Slope (mV)	E_M (mV)
Control	-23.92 ± 1.01	8.41 ± 0.89	-38.40 ± 1.50
100 µM Tannic acid	-44.95 ± 2.49 (<i>p</i> =0.0004; <i>n</i> =5)	13.67 ± 2.01 (<i>p</i> =0.0575; <i>n</i> =5)	-54.40 ± 1.23 (<0.0001; <i>n</i> =5)

Statistics versus Kv1.1 in absence of Tannic acid. Values indicate mean ± SEM.

Figure 3

	V_{0.5} Normalized tail current (mV)	Slope (mV)	E_M (mV)
Control	-21.91 ± 1.77	12.21 ± 1.58	-41.40 ± 1.00
1:50 Wild Oak Bark	-38.27 ± 1.73 (<i>p</i> =0.0002; <i>n</i> =5)	10.44 ± 1.53 (<i>p</i> =0.4442; <i>n</i> =5)	-53.40 ± 0.63 (<0.0001; <i>n</i> =5)

Statistics versus Kv1.1 in absence of White Oak Bark. Values indicate mean ± SEM.

	V_{0.5} Normalized tail current (mV)	Slope (mV)	E_M (mV)
Control	-22.59 ± 1.76	11.34 ± 1.56	-45.40 ± 1.20
1:50 Cramp Bark	-35.31 ± 1.14 (<i>p</i> =0.0006; <i>n</i> =5)	10.69 ± 0.74 (<i>p</i> =0.8283; <i>n</i> =5)	-48.20 ± 1.77 (<i>p</i> =0.4146; <i>n</i> =5)

Statistics versus Kv1.1 in absence of Cramp Bark. Values indicate mean ± SEM.

	V_{0.5} Normalized tail current (mV)	Slope (mV)	E_M (mV)
Control	-22.19 ± 3.57	17.40 ± 3.49	-49.00 ± 1.32
1:50 Wild Cherry Bark	-36.46 ± 1.19 (<i>p</i> =0.0133; <i>n</i> =5)	11.43 ± 1.06 (<i>p</i> =0.1659; <i>n</i> =5)	-58.40 ± 0.97 (<i>p</i> =0.0346; <i>n</i> =5)

Statistics versus Kv1.1 in absence of Wild Cherry Bark. Values indicate mean ± SEM.

	V_{0.5} Normalized tail current (mV)	Slope (mV)	E_M (mV)
Control	-21.46 ± 2.60	11.75 ± 2.31	-45.50 ± 0.91
1:50 White Willow Bark	-36.28 ± 0.83 (<i>p</i> =0.0032; <i>n</i> =5)	10.69 ± 0.74 (<i>p</i> =0.6810; <i>n</i> =5)	-57.40 ± 0.20 (<0.0001; <i>n</i> =5)

Statistics versus Kv1.1 in absence of White Willow Bark. Values indicate mean ± SEM.

	V_{0.5} Normalized tail current (mV)	Slope (mV)	E_M (mV)
Control	-28.46 ± 1.36	8.47 ± 1.04	-48.33 ± 0.26
1:50 <i>Sophora Japonica</i>	-45.86 ± 2.43 (<i>p</i> =0.0430; <i>n</i> =6)	8.31 ± 2.04 (<i>p</i> =0.1863; <i>n</i> =6)	-65.83 ± 2.29 (<i>p</i> =0.0002; <i>n</i> =6)

Statistics versus Kv1.1 in absence of *Sophora Japonica*. Values indicate mean ± SEM.

	V_{0.5} Normalized tail current (mV)	Slope (mV)	E_M (mV)
Control	-19.83 ± 0.65	7.93 ± 0.51	-35.12 ± 3.20
100 μM Oxymatrine	-21.86 ± 0.64 (<i>p</i> =xxx; n=8)	8.73 ± 0.24 (<i>p</i> =xxx; n=8)	-39.88 ± 2.34 (<i>p</i> =0.1720; n=8)

Statistics versus Kv1.1 in absence of Oxymatrine. Values indicate mean ± SEM.

Figure 4

KV1.1	EC50 (nM)
Tannic acid	136 ± 30 (n=7-12)
Gallic acid	379 ± 28 (n=5)
Rutin	363 ± 98 (n=5)

Kv1.1 dose responses for tannic acid, gallic acid, and rutin. Values indicate mean ± SEM.

KV1.2	EC50 (nM)
Tannic acid	222 ± 45 (n=5)
Gallic acid	<i>n.a</i> (n=5)
Rutin	855 ± 96 (n=5)

Kv1.2 dose responses for tannic acid, gallic acid, and rutin. Values indicate mean ± SEM. *n.a* = not applicable.

Figure 5

	V_{0.5} Normalized tail current (mV)	Slope (mV)	E_M (mV)
Control	-31.61 ± 1.35	5.87 ± 1.19	-43.00 ± 0.97
1:50 Bladderwrack Kelp	-46.71 ± 2.07 (<i>p</i> =0.0005; n=5)	3.99 ± 1.63 (<i>p</i> =0.3813; n=5)	-54.40 ± 0.84 (<0.0001; n=5)

Statistics versus KV1.1/KV1.1-L155P in absence of Bladderwrack Kelp. Values indicate mean ± SEM.

	V_{0.5} Normalized tail current (mV)	Slope (mV)	E_M (mV)
Control	-35.76 ± 0.92	5.88 ± 0.79	-45.80 ± 0.76
1:50 Pacific Ninebark Root	-40.80 ± 0.98 (<i>p</i> =0.0057; n=5)	5.23 ± 0.90 (<i>p</i> =0.6023; n=5)	-48.80 ± 1.23 (<i>p</i> =0.0046; n=5)

Statistics versus KV1.1/KV1.1-L155P in absence of Pacific Ninebark Root. Values indicate mean ± SEM.

	V_{0.5} Normalized tail current (mV)	Slope (mV)	E_M (mV)
Control	-22.20 ± 0.62	4.76 ± 0.55	-38.40 ± 0.31
1:50 Pacific Ninebark Leaves	-32.30 ± 0.90 (<0.0001; n=5)	6.48 ± 0.78 (p=1134; n=5)	-48.00 ± 0.23 (p=0.0036; n=5)

Statistics versus Kv1.1/Kv1.1-L155P in absence of Pacific Ninebark Leaves. Values indicate mean ± SEM.

	V_{0.5} Normalized tail current (mV)	Slope (mV)	E_M (mV)
Control	-36.99 ± 0.41	5.00 ± 0.35	-46.20 ± 0.68
1:50 Nettle	-39.21 ± 0.55 (p=0.0133; n=5)	3.88 ± 0.60 (p=0.1546; n=5)	-49.00 ± 0.20 (p=0.0348; n=5)

Statistics versus Kv1.1/Kv1.1-L155P in absence of Nettle. Values indicate mean ± SEM.

Figure 6

	V_{0.5} Normalized tail current (mV)	Slope (mV)	E_M (mV)
Control	-11.63 ± 1.11 (n=5)	7.55 ± 2.83 (n=5)	-30.80 ± 0.43
1:50 Bladderwrack Kelp	-29.20 ± 0.65 (<0.0001; n=5)	6.52 ± 0.80 (p=0.7415; n=5)	-47.80 ± 0.84 (p=0.0025; n=5)

Statistics versus Kv1.1-E283K in absence of bladderwrack kelp. Values indicate mean ± SEM.

	V_{0.5} Normalized tail current (mV)	Slope (mV)	E_M (mV)
Control	-4.98 ± 0.47 (n=5)	4.14 ± 1.59 (n=5)	-22.00 ± 0.28
1:50 Pacific Ninebark Root	-19.24 ± 0.87 (<0.0001; n=5)	3.27 ± 1.63 (p=0.7124; n=5)	-34.20 ± 0.85 (p=0.0009; n=5)

Statistics versus Kv1.1-E283K in absence of Pacific Ninebark root. Values indicate mean ± SEM.

	V_{0.5} Normalized tail current (mV)	Slope (mV)	E_M (mV)
Control	-7.80 ± 1.03 (n=5)	7.03 ± 3.93 (n=5)	-24.80 ± 1.24
1:50 Pacific Ninebark Leaves	-19.88 ± 0.76 (<0.0001; n=5)	6.05 ± 2.81 (p=0.8448; n=5)	-35.00 ± 0.84 (p=0.0003; n=5)

Statistics versus Kv1.1-E283K in absence of Pacific Ninebark Leaves. Values indicate mean ± SEM.

	V_{0.5} Normalized tail current (mV)	Slope (mV)	E_M (mV)
Control	-5.84 ± 1.09 (n=6)	6.18 ± 0.95 (n=6)	-35.40 ± 0.74
1:50 Nettle	-24.17 ± 0.55 (<0.0001; n=6)	5.00 ± 0.47 (p=0.2268; n=6)	-47.00 ± 0.83 (<0.0001; n=6)

Statistics versus Kv1.1-E283K in absence of Nettles. Values indicate mean ± SEM.

	Control Tau_{Act} (ms)	1 μM Tannic acid Tau_{Act} (ms)
-20 mV	19.99 \pm 7.47	41.32 \pm 8.47 (<i>p</i> =0.0962; n=5)
-10 mV	10.66 \pm 4.72	27.06 \pm 7.08 (<i>p</i> =0.0955; n=5)
0 mV	8.72 \pm 4.64	21.12 \pm 4.35 (<i>p</i> =0.0872; n=5)
+10 mV	4.26 \pm 1.26	17.43 \pm 3.97 (<i>p</i> =0.0265; n=5)
+20 mV	3.86 \pm 1.20	12.72 \pm 2.85 (<i>p</i> =0.0324; n=5)
+30 mV	3.40 \pm 1.06	10.63 \pm 2.79 (<i>p</i> =0.0587; n=5)
+40 mV	3.09 \pm 0.91	10.04 \pm 2.65 (<i>p</i> =0.0565; n=5)

Statistics versus Kv1.1-E283K in absence of bladderwrack kelp. Values indicate mean \pm SEM.

	Control Tau_{Deact} (ms)	1 μM Tannic acid Tau_{Deact} (ms)
-80 mV	0.88 \pm 0.32	1.50 \pm 0.57 (<i>p</i> =0.3632; n=8)
-70 mV	0.38 \pm 0.15	1.38 \pm 0.56 (<i>p</i> =0.1228; n=8)
-60 mV	0.32 \pm 0.09	1.23 \pm 0.52 (<i>p</i> =0.1259; n=8)
-50 mV	0.20 \pm 0.05	1.04 \pm 0.44 (<i>p</i> =0.0986; n=8)
-40 mV	0.19 \pm 0.05	0.88 \pm 0.36 (<i>p</i> =0.0979; n=8)
-30 mV	0.19 \pm 0.05	0.79 \pm 0.32 (<i>p</i> =0.1044; n=8)
-20 mV	0.19 \pm 0.06	0.87 \pm 0.36 (<i>p</i> =0.1025; n=8)

Statistics versus Kv1.1-E283K in absence of bladderwrack kelp. Values indicate mean \pm SEM.

Figure 7

	V_{0.5} Normalized tail current (mV)	Slope (mV)	E_M (mV)
Control	<i>n.a</i>	<i>n.a</i>	-41.20 ± 1.84
1 μM Gallic acid	<i>n.a</i>	<i>n.a</i>	-47.00 ± 2.56 (<i>p</i> =0.1067; <i>n</i> =5)

Statistics versus Kv1.1/Kv1.1-G311D in absence of Gallic acid. Values indicate mean ± SEM. *n.a* = not applicable.

	V_{0.5} Normalized tail current (mV)	Slope (mV)	E_M (mV)
Control	<i>n.a</i>	<i>n.a</i>	-25.40 ± 1.37
1 μM Gallic acid	<i>n.a</i>	<i>n.a</i>	-33.00 ± 1.09 (<i>p</i> =0.0131; <i>n</i> =5)

Statistics versus Kv1.1/Kv1.1-L328V in absence of Gallic acid. Values indicate mean ± SEM. *n.a* = not applicable.

	V_{0.5} Normalized tail current (mV)	Slope (mV)	E_M (mV)
Control	<i>n.a</i>	<i>n.a</i>	-41.40 ± 1.98
1 μM Gallic acid	<i>n.a</i>	<i>n.a</i>	-46.60 ± 0.60 (<i>p</i> =0.0289; <i>n</i> =5)

Statistics versus Kv1.1/Kv1.1-V408A in absence of Gallic acid. Values indicate mean ± SEM. *n.a* = not applicable.

	V_{0.5} Normalized tail current (mV)	Slope (mV)	E_M (mV)
Control	-12.01 ± 0.57	4.52 ± 0.51	-33.80 ± 0.18
1 μM Gallic acid	-27.50 ± 0.51 (<0.0001; <i>n</i> =4)	5.70 ± 1.12 (<i>p</i> =0.3772; <i>n</i> =4)	-47.80 ± 0.65 (<i>p</i> =0.0005; <i>n</i> =4)

Statistics versus Kv1.1/Kv1.1-E283K in absence of Gallic acid. Values indicate mean ± SEM.

Figure 8

	V_{0.5} Normalized tail current (mV)	Slope (mV)	E_M (mV)
Control	-23.84 ± 0.45	4.30 ± 0.35	-34.89 ± 0.98 (n=9)
1 µM Gallic acid	-30.59 ± 0.60 (<0.0001; n=9)	4.84 ± 0.56 (p=0.4278; n=9)	-45.00 ± 0.78 (p=0.0002; n=9)
10 µM Gallic acid	-36.38 ± 0.62 (<0.0001; n=9)	4.76 ± 0.51 (p=0.4692; n=9)	-48.89 ± 0.66 (<0.0001; n=9)
100 µM Gallic acid	-37.93 ± 0.62 (<0.0001; n=9)	4.68 ± 0.55 (p=0.5695; n=9)	-50.11 ± 0.69 (<0.0001; n=9)

Statistics versus Kv1.1/Kv1.1-L155P in absence of Gallic acid. Values indicate mean ± SEM.

Figure 9

	V_{0.5} Normalized tail current (mV)	Slope (mV)	E_M (mV)	Current-fold change -30 mV
Control	-11.21 ± 0.61	5.13 ± 0.60	-27.00 ± 0.35	<i>n.a</i>
1 µM Tannic acid	-22.75 ± 1.34 (<0.0001; n=6)	8.70 ± 1.19 (p=0.0301; n=6)	-44.83 ± 0.14 (<0.0001; n=6)	4.69 ± 1.69 (<0.0001; n=6)

Statistics versus Kv1.1-E283K in absence of Tannic acid. Values indicate mean ± SEM. *n.a* = not applicable.

	Control Tau_{Act} (ms)	1 µM Tannic acid Tau_{Act} (ms)
-20 mV	24.46 ± 3.97	31.31 ± 2.98 (p=0.2679; n=6)
-10 mV	15.41 ± 2.70	20.71 ± 2.35 (p=0.1701; n=6)
0 mV	11.02 ± 1.62	15.70 ± 1.40 (p=0.0543; n=6)
+10 mV	8.90 ± 0.89	12.94 ± 1.05 (p=0.0153; n=6)
+20 mV	7.43 ± 0.68	11.30 ± 0.89 (p=0.0068; n=6)
+30 mV	6.35 ± 0.66	10.16 ± 0.83 (p=0.0053; n=6)
+40 mV	5.50 ± 0.69	9.46 ± 0.83 (p=0.0046; n=6)

Statistics versus Kv1.1-E283K in absence of Tannic acid. Values indicate mean ± SEM.

	Peak current at -40 mV (μ A)
Control	0.02 \pm 0.01
0.1 μ M	0.02 \pm 0.01 (>0.9999 ; n=5)
1 μ M	0.02 \pm 0.01 (>0.9999 ; n=5)
10 μ M	0.02 \pm 0.01 (>0.9999 ; n=5)
30 μ M	0.02 \pm 0.01 (>0.9999 ; n=5)
100 μ M	0.03 \pm 0.01 ($p=0.496$; n=5)

Statistics versus Kv1.1-E283K in absence of Tannic acid. Values indicate mean \pm SEM.

Figure 10

	EC50 (nM)
Kv1.1	379 \pm 28 (n=5)
Kv1.1-3M	<i>n.a</i>

Kv1.1 vs Kv1.1-3M dose responses for gallic acid. Values indicate mean \pm SEM. *n.a* = not applicable.

	EC50 (nM)
Kv1.1	18 \pm 6 (n=5)
Kv1.1-3M	345 \pm 38 (n=5)

Kv1.1 vs Kv1.1-3M resting membrane potential (E_M) dose responses for gallic acid. Values indicate mean \pm SEM.